

# Isolation of bovine and human milk-derived exosomes: optimization of methods



uOttawa

Andrea Zukowski<sup>1</sup>, Jamie Kraft<sup>2</sup>, Illimar Altosaar<sup>2</sup>

<sup>1</sup>Faculty of Health Sciences, University of Ottawa

<sup>2</sup>Faculty of Medicine, University of Ottawa, Department of Biochemistry, Microbiology, and Immunology

## Introduction

- Released by intact mammary cells, exosomes are extracellular microvesicles (EMs) (30-100 nm diameter)<sup>1</sup> by which a mother may communicate important signals for the infant's developing gastrointestinal epithelial cells<sup>2</sup>. Exosomes can potentially be used to prevent and treat various neonatal gastroenterology disorders, but have yet to be fully understood.
- Due to the lack of optimized protocols for, and difficulty of, isolating exosomes from other constituents present in breast and bovine milk<sup>3,4</sup>, research performed on exosomes may not be selective for these vesicles.
- Aim: to compare purification protocols for optimized yield and quality of exosomes. Both human and bovine milk were analyzed for optimal exosome isolation.

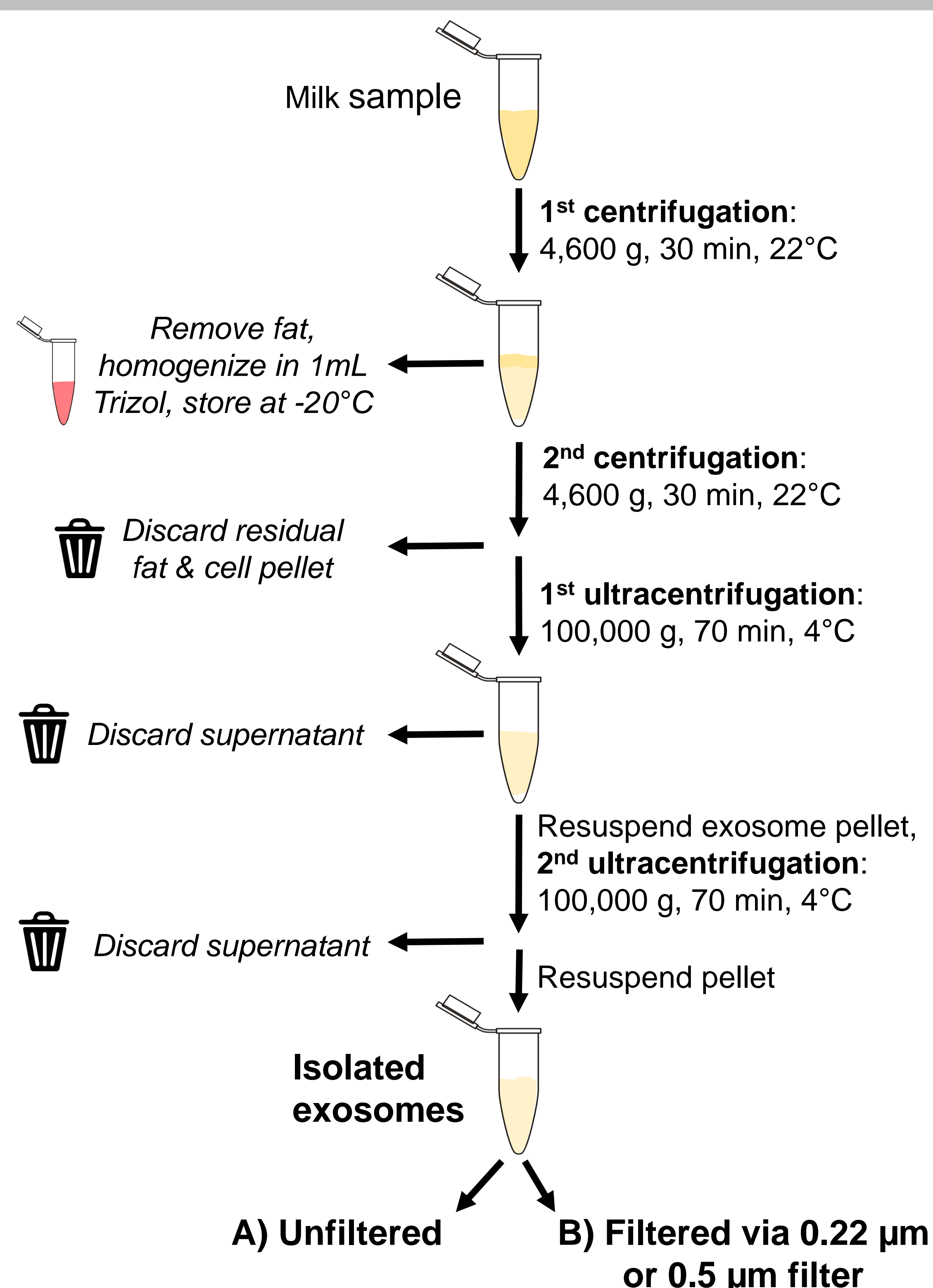
## Methodology

### Isolation:

- Exosome populations were derived from either a sample of bovine milk (whole milk, Sheldon Creek Dairy) or human breast milk (frozen at -80°C after extraction).
- All steps were performed in DEPC-treated water.
- The samples were prepared with the protocol in **Figure 1**.

### Detection:

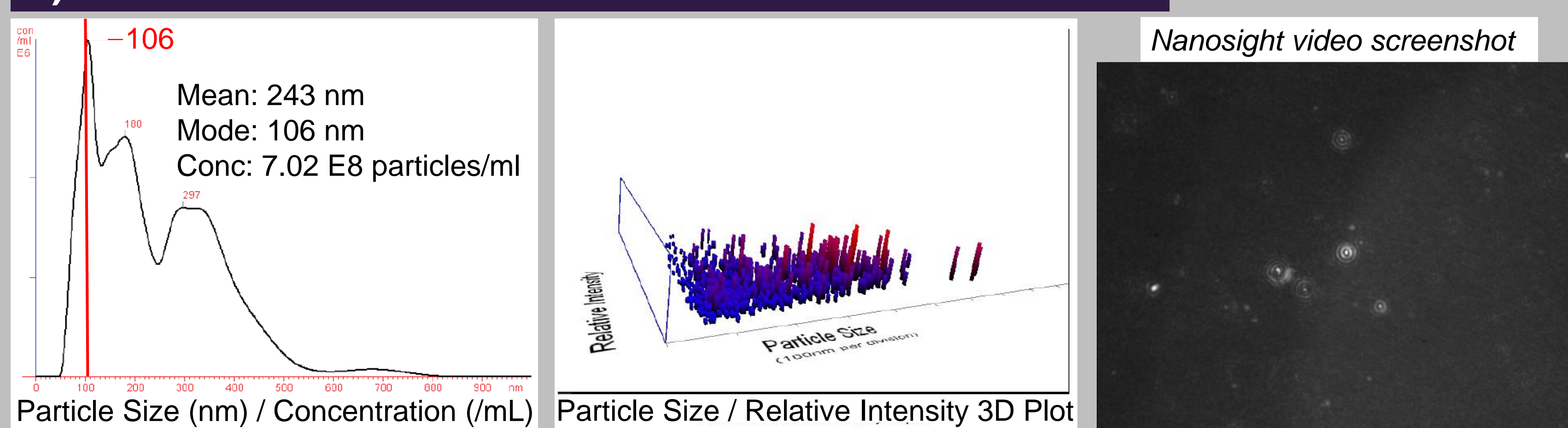
- Nanoparticle tracking analysis (Nanosight): detect the presence & concentration of vesicles based on size.
- Western blot: detect commonly found tetraspanins in exosomes, such as CD63 and CD81 receptors<sup>1</sup>.



**Figure 1.** Protocol for exosome isolation

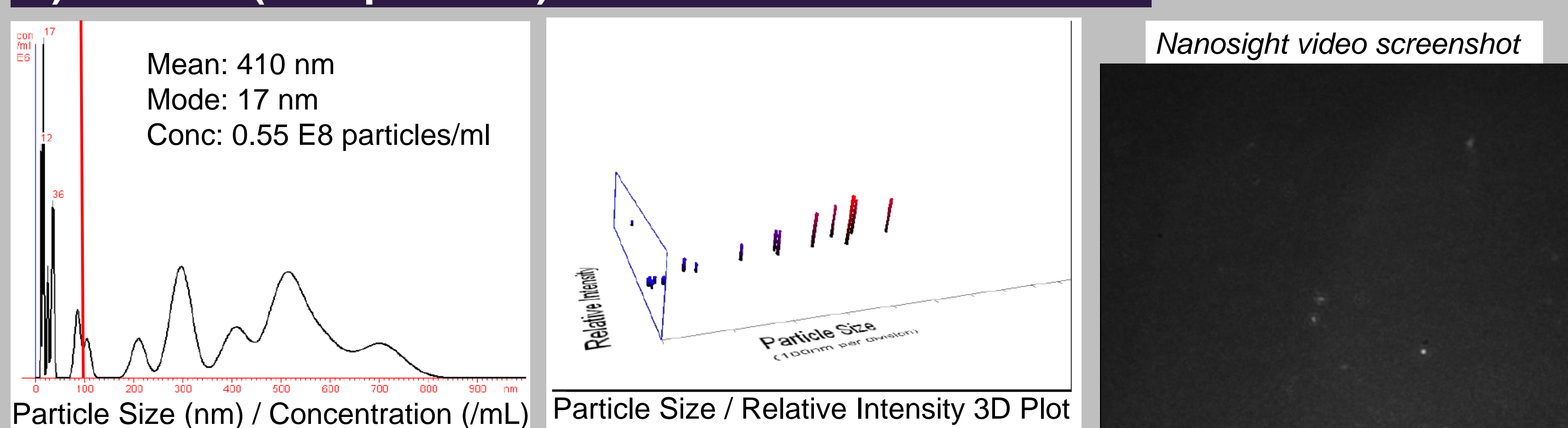
## Results

### A) Unfiltered – Breast Milk

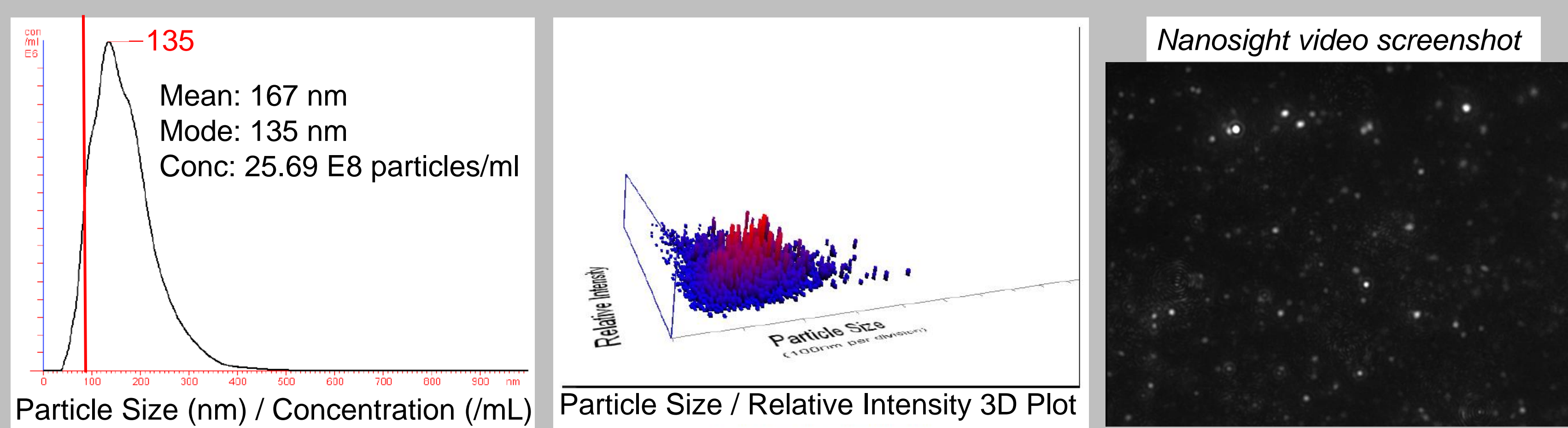


**Figure 2.** Isolated breast milk exosomes (1/10 dilution), unfiltered

### B) Filtered (0.22 µm filter) – Breast Milk

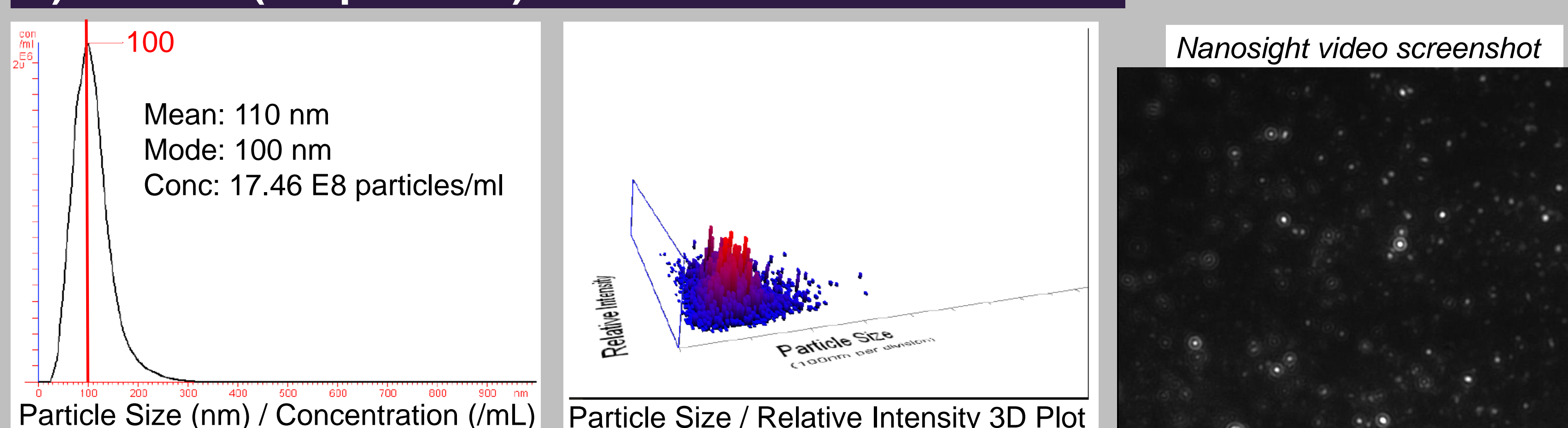


**Figure 3.** Isolated breast milk exosomes, filtered 1x with a 0.22 µm filter

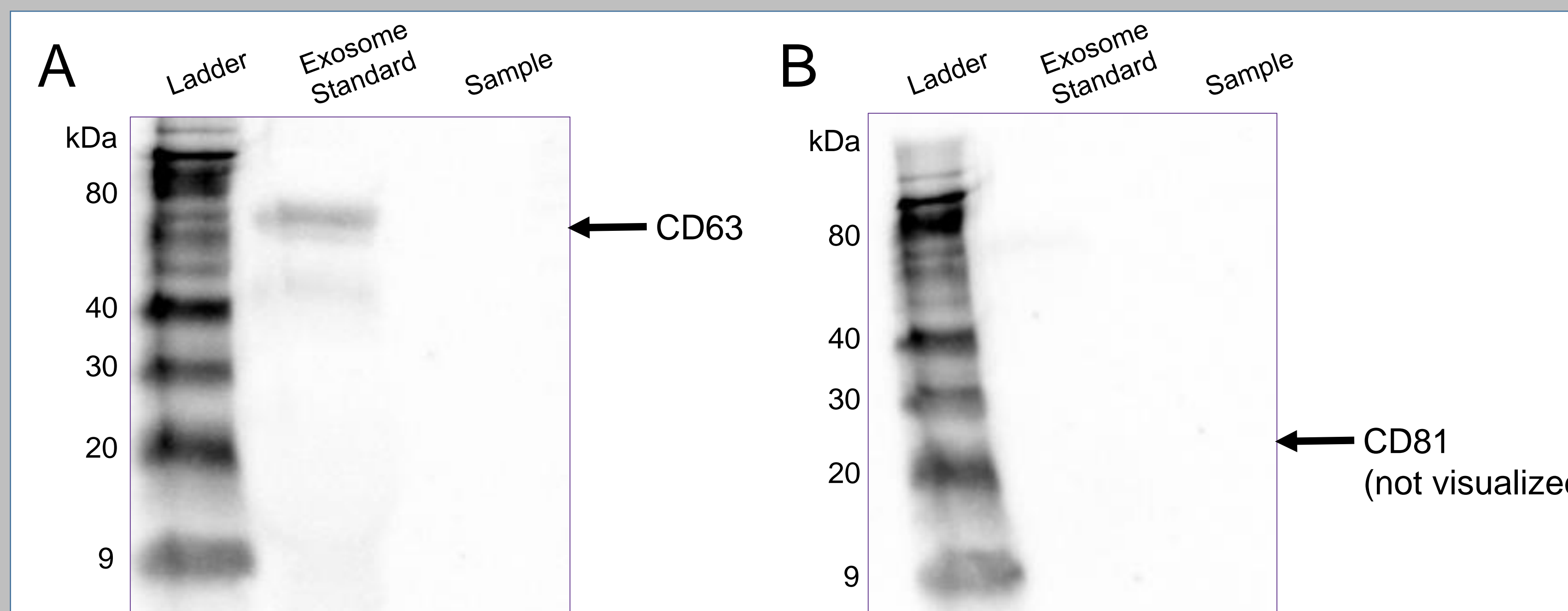


**Figure 4.** Isolated breast milk exosomes, filtered 1x with a 0.22 µm filter and 3x with a fresh 0.22 µm filter.

### B) Filtered (0.5 µm filter) – Bovine Milk



**Figure 5.** Isolated bovine milk exosomes, filtered 2x with two fresh 0.5 µm filters (1/320 dilution)



**Figure 4.** Western blot of bovine milk-derived exosomes. (A) Immunoblot to detect CD63 membrane protein. (B) Immunoblot to detect CD81 membrane protein.

## Discussion & Conclusion

- The protocol used in this study demonstrated the ability to isolate exosome-sized EMs.
- Filtration using a 0.22 µm filter (**Figure 4**) or 0.5 µm filter (**Figure 5**) removed more debris and more exosome-sized EMs. Washing and diluting the sample enabled clearer imaging.
- Vesicles isolated could not be verified to contain exosomes via Western blot, suggesting presence of other EMs, or low concentrations of exosomes in the sample.
- Further optimization of exosome isolation will help to better study the characteristics of these EMs, including their potentially immunomodulating miRNA and protein contents.

### Future directions:

- Performing the protocol in PBS to maintain membrane integrity better than DEPC-treated water, increasing the concentration of exosomes.
- Comparing results to sucrose density gradient centrifugation, which has been used to isolate exosomes<sup>4</sup>.
- Further investigation of the capability of filtration for exosome purification.
- Use of flow cytometry and electron microscopy to obtain more precise evidence of the presence of exosomes<sup>1</sup>.

### References:

- Lässer, C., et al. (2011). Human saliva, plasma and breast milk exosomes contain RNA: uptake by macrophages. *Journal of translational medicine*, 9(1), 9.
- Floris, I., Kraft, J.D., Altosaar, I. "Roles of MicroRNA across Prenatal and Postnatal Periods." *International Journal of Molecular Sciences* 17.12 (2016): 1994.
- Chen, T., et al. (2014). Exploration of microRNAs in porcine milk exosomes. *BMC genomics*, 15(1), 100.
- Zonneveld, M. I., et al. (2014). Recovery of extracellular vesicles from human breast milk is influenced by sample collection and vesicle isolation procedures. *Journal of extracellular vesicles*, 3.

## Acknowledgements

Dr. J. Siggers helped develop protocols. The Undergraduate Research Opportunity Program (UROP), CIHR and NSERC helped fund this project.

## Contact Information

Personal  
Andrea Zukowski  
Phone: (416) 876-8003  
Email: azuko037@uottawa.ca

Laboratory  
Dr. Illimar Altosaar  
Phone: (613) 562-5800 (x 6371)  
Email: altosaar@uottawa.ca