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SUBSTITUTION OF TRYPTOPHAN WITH ALANINE IN HUMAN D1 DOPAMINE RECEPTORS

TIBERI M., ZHANG B., SABIRI. P

UNIVERSITY OF OTTAWA FACULTY OF MEDICINE AND OTTAWA HOSPITAL RESEARCH INSTITUTE

Introduction^{1,2}

The key brain neurotransmitter that is involved with learning, memory, pleasure, movement and proper cognition is dopamine. It has been long understood that the improper regulation of dopamine results in complications that can lead to the development of schizophrenia, parkinson's, and drug addiction. Dopamine exerts its effects by binding to ligand membrane receptors that belong to either the hD1 or hD5 family. These receptors are transmembrane receptors with seven protein domains and three intracellular loops. The purpose of this investigation is to examine the role of certain amino acids containing hydrogen donating groups in hD1 to learn about how they interact inside the cell, and whether one can alter the function of hD1 by simply substituting these amino acids with other amino acids with a less capability to hydrogen bond. In other words, missense mutations are induced to substitute certain amino acids in the hD1 receptor and then the mutated hD1 receptors are expressed to monitor their proper functioning. The amino acid of interest in this project is tryptophan (in position 148 of protein), an indole containing amino acid that can hydrogen bond by donating hydrogens. It will be substituted with alanine, a non-indole amino acid with a lower hydrogen bonding capability. In the past, many experiments have been conducted to explain the role of intracellular loop three (IL3) in the functioning of hD1 and hD5, but still not enough research has shed light onto this subject. Interestingly, not much is known about how dopamine receptors function. Hopefully, this investigation will enlighten current knowledge about dopamine receptors and will allow further investigations to develop a complete three dimensional model of these receptors. Having a 3D model allows for the development of specific therapeutic drugs that can potentially treat the neurological disorders mentioned above with minimal side effects.

Methods

A molecular biology approach was taken to create the desired amino acid substitution.

- First, PCR amplifications were used to generate megaprimers and the overlap
- Second, a DNA digestion was carried out using the enzymes EcoRI and HindIII to isolate the desired cassette
- Third, DNA ligation was conducted to link the cassette to the vector
- Fourth, XL1-blue bacteria were transformed with the newly made vector through electroporation
- Finally, bacterial colonies were isolated and a DNA extract was collected for sequencing

View figures 1 and 2.

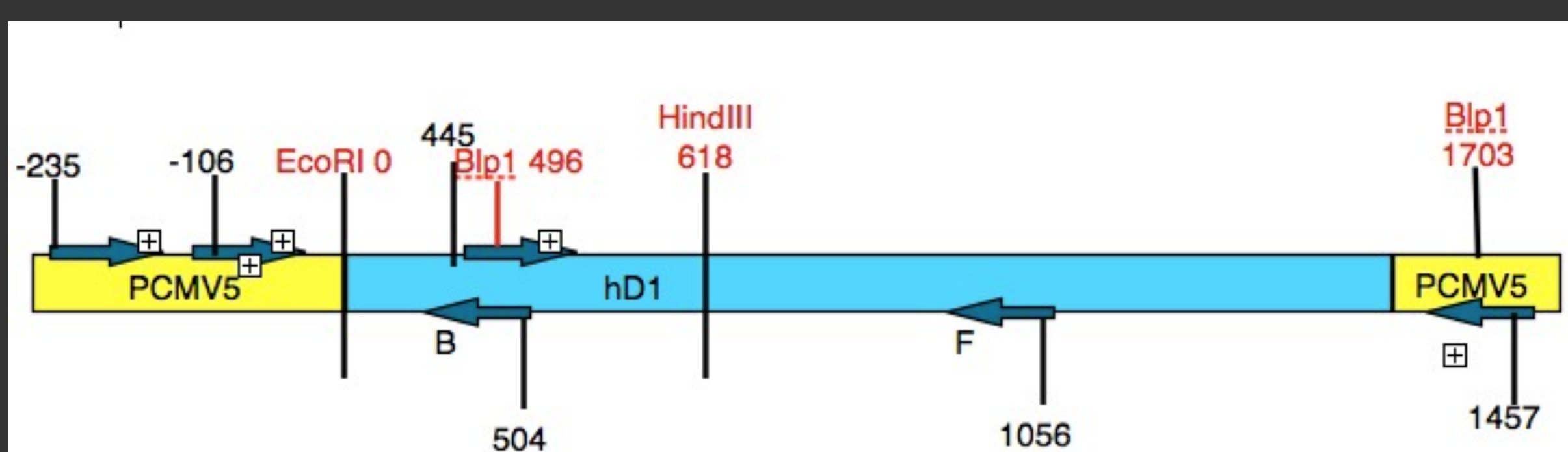


Figure 1: hD1 gene showing the binding sites of primers and enzyme cut sites for generation of the megaprimers and then the overlap.

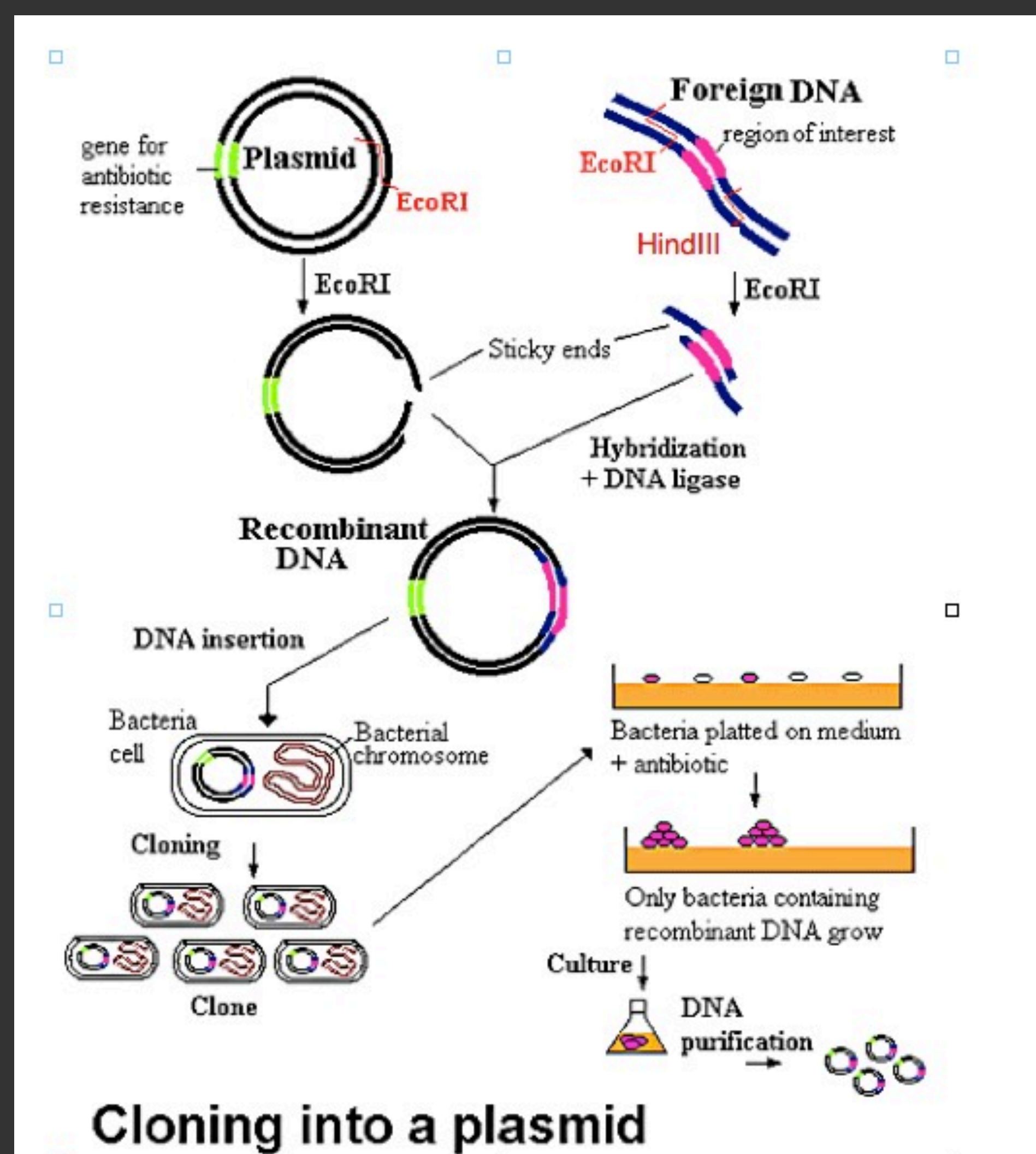


Figure 23: The insertion of a desired sequence into a plasmid and then the transformation of the plasmid into the XL1-blue bacteria.

Results

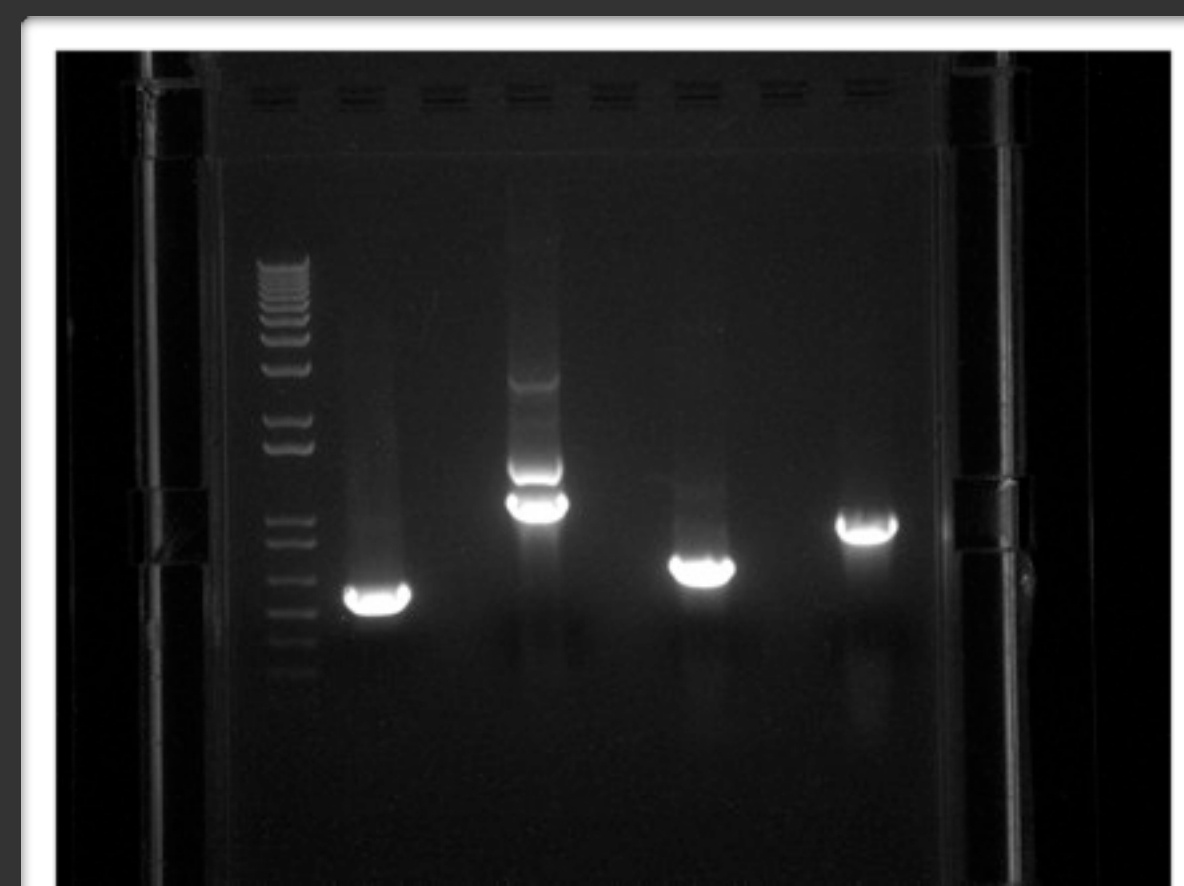


Figure 3: Electrophoresis gel showing megaprimers AB and CD

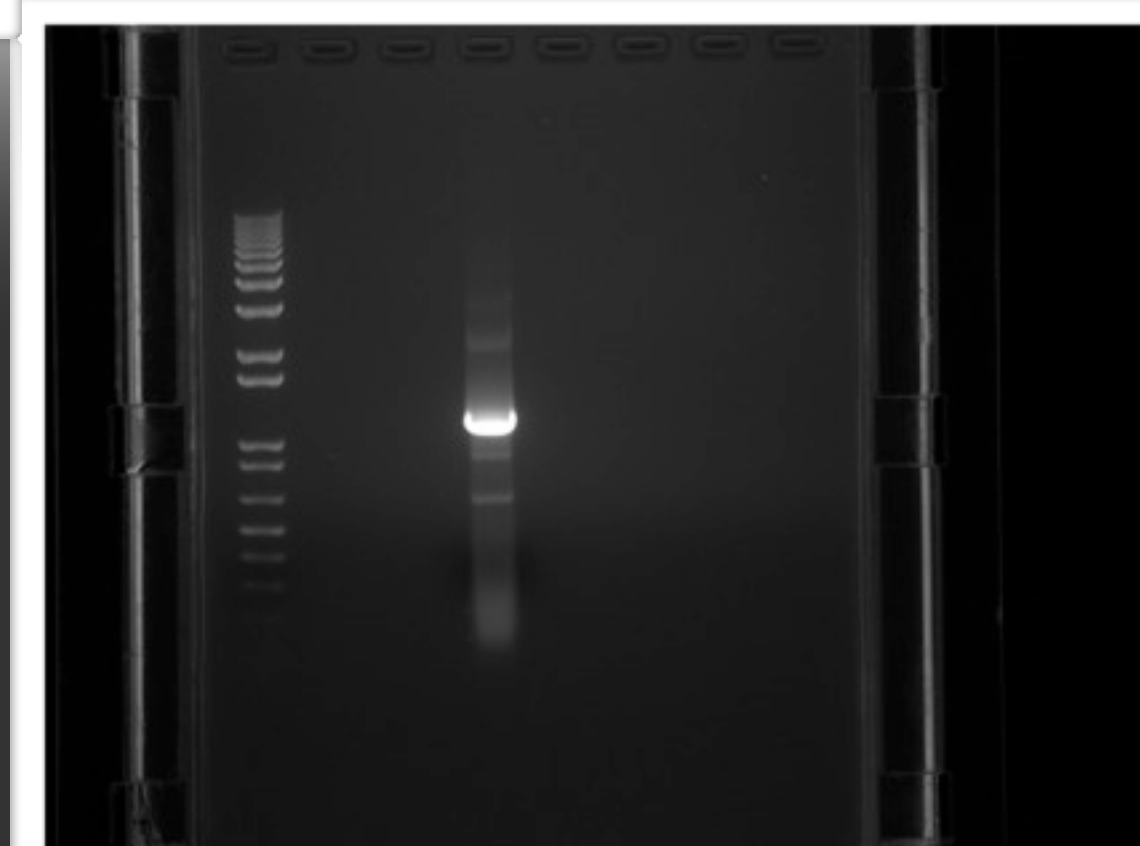


Figure 4: Electrophoresis image showing DNA overlap.

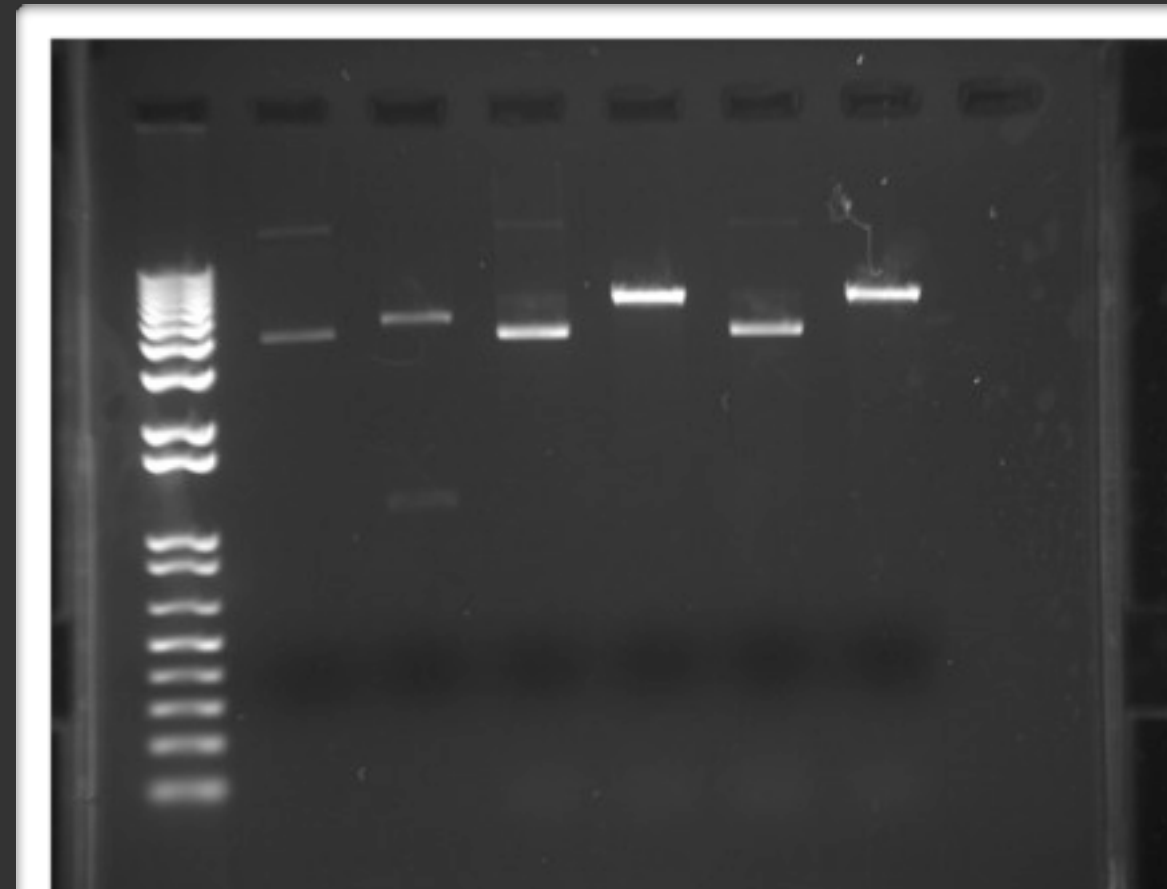


Figure 5: Electrophoresis image showing the cassette.

hD1-W148A
 tacgggtgggaggtctatataaacagagagctcgttttagtgaaccctcagaaattcggatccaagATG
 AGGACTCTGAACACCTCTGCCATGGACGGGACTGGGCTGGTGGGAGAGGACTTCTCTGTTC
 GTATCCTCACTGCCTGTTTCTCGCTGCTCATCTCCCTGCCAGCTCTGGGGAACACGCTGGT
 CTGTGCTGCCGTTATCAGGTTCCGACACCTGGGTCACAGGTGACCACTTCTTTGTCATCTCC
 TTGGCTGTGCAGATCTCTTGGTGGCCCTCCCTGGTCACTGCTGGAGGCGAGTGGCTGAGATG
 CTGGCTCTGGCCCTTGGGCTCTCTGTAAACATCTGGGTCCTTTGACATGATGCTGCTCAC
 TGCATCCATCCCTCAAGCTCTGTGTATCAGGCTGGACAGGTATGGGCTATCTCCAGCCCTTC
 CGGTATGAGAGAAATGACCCCAAGGCGCTTCACTCCATCATGCTGTGGCAGCCACTTGT
 CTGTACTCATCTCTTATCCAGTGAACCTCAGCTGGCAGCAGGCAAAACCCCAAGCCCTC
 TGATGAAATGCCACTTCCCTGGCTGAGACCATAGACAACTGTGACTCCAGCTCAGCAGGACA
 TATGCCATCTCATCTCTGTAAAGCTTTACATCCCTGTGGCCATCATGATTGTCACTACA
 CCAGGATCTACAGGATTGCTCAGAACAAATACGGCCATTCGGCCCTTGAGAGGGCGAGCAT
 CCACCCCAAGAATTGCCAGACCCACAGGTAAATGAAAGCCCTGCGAATGTTCTCAACCCGAA
 AGTCTTTAAGATGCTCTCAAGAGCAACTAAAGCTCTGAGAGCTCTGTGGTGAATG
 GTGTCTTGTGCTGCTTGGCTCTCTCTCACTTGAACGCAATTTGCCCTCTGTGGTTC
 TGGGAGACCGCCCTTCTGCATTGATCCAAACCTTTGACGTTTGTGGTGGTGGTGG
 GCTAATCATCTTGAACCCATATTTATGCTTTAATGCTGATTTTCGGAAGGCATTTTCAA
 CCCTTAGGATGCTACAGACTTGGCCCTGGCAGCAATAATGCCATAGACGCTGAGATCAA
 TAACAATGGGCGCGATGTTTCCAGCCATCATGAGCCAGGCTCCATCTCCAAGAGTGC
 AATCTGTTTACCTGATCCACATGCTGTGGCTCCTCTGAGGACCTGAAAAGAGGAGGAGCAG
 CTGGCATCCGACCCCTGGAGAGCTGTCCCGACCCCTATCGGTCATATGGACTATGACAC
 TGAGCTCTCTGGAGAGATCCACCCATCACAAACGGTCCAGCACCACCTGAGGATcc
 tctaaagagatccgggtggcatccctgtgacccctcccaagtgccctctccctggccctggaagt
 gcoactcagtgccaccagcctgtctcaataaaatata

Legend:
 Blue = enzyme sequence cutting the cassette
 Red = Mutations for generating alanine
 Orange = Silent mutation used for screening

Figure 64: DNA Sequence of hD1 gene showing missense mutation

Upon sequencing the DNA in the bacterial extract, the desired missense mutation was found. Bacterial colonies had grown on a medium containing nutrients and an antibiotic.

Figures 3,4, and 5 show the gel electrophoresis of the megaprimers, the overlap, and the cassette respectively. Figure 6 shows the DNA sequence of the cassette after it was isolated from the bacterial colonies along with a legend. Figure 3 indicates that there is a megaprimer around 739 base pairs (bp) and another megaprimer around 1012 bp. Figure 4 shows the overlap product at around 1162 bp. Finally, figure 6 shows the DNA sequence of the cassette and a GC base mutation within that sequence.

Conclusion

In conclusion, the hD1 gene has been mutated to generate alanine instead of tryptophan in the hD1 human receptor. The DNA sequence obtained from the bacterial extract shows that the bases G and C were incorporated in the DNA. The mutated gene can now be inserted in human basal cells where the dopamine receptor will be expressed. Further investigations will be able to measure the functioning of the receptor and how it interacts with drugs, hopefully shedding light about the structural and functional characteristics of the different regions of hD1 receptor.

Bibliography

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